

AD-A168 819

PERIPHERAL NERVE INJURY IN DEVELOPING RATS REORGANIZES  
MOTOR CORTEX(U) BROWN UNIV PROVIDENCE RI CENTER FOR  
NEURAL SCIENCE J P DONOGHUE ET AL. 19 MAY 86 TR-29

1/1

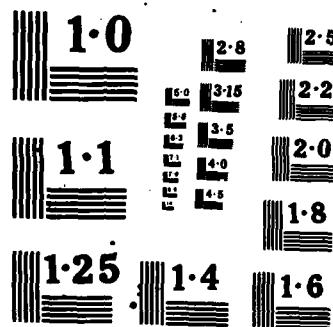
UNCLASSIFIED

N00014-81-K-0136

F/G 6/5

NL





NATIONAL BUREAU OF S  
MONOCOPY RESOLU TEST

13

REPORT DOCUMENTATION PAGE		READ INSTRUCTIONS BEFORE COMPLETING FORM
REPORT NUMBER #29	2. GOVT ACCESSION NO.	3. RECIPIENT'S CATALOG NUMBER
TITLE (and Subtitle) Peripheral Nerve Injury in Developing Rats Reorganizes Motor Cortex		5. TYPE OF REPORT & PERIOD COVERED Technical Report
AUTHOR(s) John P. Donoghue and Jerome N. Sanes		6. PERFORMING ORG. REPORT NUMBER
PERFORMING ORGANIZATION NAME AND ADDRESS Center for Neural Science Brown University Providence, Rhode Island 02912		8. CONTRACT OR GRANT NUMBER(s) N00014-81-K-0136, #PHS-1-K-1-00014-01
11. CONTROLLING OFFICE NAME AND ADDRESS Personnel and Training Research Program Office of Naval Research, Code 442PT Arlington, Virginia 22217		10. PROGRAM ELEMENT, PROJECT, TASK AREA & WORK UNIT NUMBERS N-201-484
14. MONITORING AGENCY NAME & ADDRESS (if different from Controlling Office)		12. REPORT DATE May 19, 1986
		13. NUMBER OF PAGES 12 pages
		15. SECURITY CLASS. (of this report) Unclassified
		15a. DECLASSIFICATION/DOWNGRADING SCHEDULE
16. DISTRIBUTION STATEMENT (of this Report) Approved for public release; distribution unlimited. Publication in part or in whole is permitted for any purpose of the United States Government.		
17. DISTRIBUTION STATEMENT (of the abstract entered in Block 20, if different from Report)		
18. SUPPLEMENTARY NOTES  <b>DTIC ELECTED</b> <b>S</b> MAY 27 1986 <b>A</b>		
19. KEY WORDS (Continue on reverse side if necessary and identify by block number) motor cortex neuroplasticity development cerebral cortex		
20. ABSTRACT (Continue on reverse side if necessary and identify by block number) We investigated the effect of neonatal nerve lesions on cerebral motor cortex organization by comparing the cortical motor representation of normal adult rats with adult rats that had one forelimb removed on the day of birth. Mapping of cerebral neocortex with electrical simulation revealed an altered relationship between the motor cortex and the remaining muscles. While distal forelimb movements are normally elicited at the lowest threshold in the motor cortex forelimb area, the same stimuli activated shoulder and trunk muscles in experimental animals. In addition an expanded cortical representation of intact body		

DD FORM 1 JAN 73 1473 EDITION OF 1 NOV 68 IS OBSOLETE  
S/N 0102-LF-014-6601

SECURITY CLASSIFICATION OF THIS PAGE (When Data Entered)

86 5 27 004

cont'd

parts was present and there was an absence of a distinct portion of motor cortex. These data demonstrate that representation patterns in motor cortex are subject to environmental events that occur during development. *Key words*

*Neuroplasticity*

Accession For	
NTIS GRA&I	<input checked="" type="checkbox"/>
DTIC TAB	<input type="checkbox"/>
Unannounced	<input type="checkbox"/>
Justification	
By _____	
Distribution/ _____	
Availability Codes	
Dist	Avail and/or Special
A1	



S-N 0102-LF-014-6601

Peripheral Nerve Injury in Developing Rats Reorganizes  
Motor Cortex

John P. Donoghue

Center for Neural Science

Brown University

Providence, Rhode Island 02912

and

Jerome N. Sanes

Human Motor Control Section

Medical Neurology Branch

National Institute of Neurological and Communicative Disorders and Stroke

Bethesda, MD 20892

---

Supported in part by the U. S. Office of Naval  
Research Contract #N00014-81-K-0136

*Abstract. We investigated the effect of neonatal nerve lesions on cerebral motor cortex organization by comparing the cortical motor representation of normal adult rats with adult rats that had one forelimb removed on the day of birth. Mapping of cerebral neocortex with electrical stimulation revealed an altered relationship between the motor cortex and the remaining muscles. While distal forelimb movements are normally elicited at the lowest threshold in the motor cortex forelimb area, the same stimuli activated shoulder and trunk muscles in experimental animals. In addition an expanded cortical representation of intact body parts was present and there was an absence of a distinct portion of motor cortex. These data demonstrate that representation patterns in motor cortex are subject to environmental events that occur during development.*

Among all cerebral neocortical areas, the motor cortex (MI) appears to be most closely related to the control of muscles. MI is necessary for the independent use of muscles in skilled voluntary movements, participates in movement initiation and is involved in the elaboration of the complex repertoire of movements of mammals (1). Genetic mechanisms expressed during development are believed to be important determinants of the functional organization of cortical areas. Nevertheless, it is clear that environmental events modify representational patterns, at least in sensory cortex. Anatomical and electrophysiological investigations of somatic sensory and visual cortices have shown that injury to peripheral receptors or even selective forms of experience in developing mammals results in expanded representations of intact parts (2) and concomitant changes in cortical connectivity patterns (3-5). Although developmentally induced modifications in representation patterns, intrinsic organization or efferent connections of motor cortex could profoundly affect movement execution and control, the effect of peripheral environmental factors on these aspects of MI organization is largely unknown.

MI is defined in a variety of mammals as the cortical area in which movements can be evoked at the lowest levels of electrical stimulation (6). Intracortical microstimulation (ICMS) mapping reveals a topographically ordered pattern of representation in MI; focal sites in cortex are related to one or a few closely related muscles (7). In rats MI is largely comprised of a cytoarchitectonically distinct frontal agranular cortical field that contains forelimb, head and trunk representations. A portion of the immediately adjacent granular somatic sensory cortex (SI) is also included in MI because ICMS in this region evokes movements at thresholds comparable to those in the agranular part of MI (8-10). This amalgamation of SI and MI, called the overlap zone (OLZ), is composed of the motor and somatic sensory representations of the distal forelimb and hindlimb (9, 11).

In the present experiments we examined the effects of nerve lesions on the somatic representation pattern in MI. The organization of MI was studied with ICMS in 11 normal adult animals and in 14 adult rats that had the right forelimb amputated on the day of birth (12). The experimental animals were allowed to recover for 8-24 weeks whereupon MI and the adjacent

cortex were mapped with ICMS or extracellular recording techniques (13). We first defined the forelimb representation in the agranular and OLZ regions of MI in normal animals and then mapped a similar cortical location in the experimental animals. At each site of electrode penetration the body part that was activated at the lowest current intensity was identified by a combination of visual inspection and muscle palpation; other body parts activated at higher currents up to 60  $\mu$ A were noted.

The low-threshold cortical maps generated from one normal and one experimental rat are shown in Fig. 1A and 1B. In intact rats the MI forelimb area is typically separated into distinct caudal and rostral regions (14). The larger caudal region comprises contiguous areas in the agranular part of MI and in the OLZ. The rostral part of the forelimb region is located exclusively in agranular cortex. In the normal rat, threshold ICMS within the MI forelimb area only activates muscles of the contralateral distal forelimb, producing movements of the digits, wrist or elbow. Muscles of proximal body parts are activated at some distal forelimb sites, but only at higher current intensities. We never observed a clear case of shoulder muscle activation at the lowest current intensity, although at one site the neck and shoulder muscles appeared to have the same threshold. We assume that the higher threshold for proximal movements represents either less effective synaptic connections or a greater number of relays between cortical sites and the proximal muscles. The possibility that the higher currents required to activate proximal limb muscles signify current spread is ruled out by the consistent finding that nowhere in MI of normal rats can a separate low threshold shoulder zone be found.

In experimental animals, we defined the MI forelimb region as that area in which ICMS activated muscles about the contralateral shoulder or ipsilateral forelimb. This region was smaller than that of normal rats (normal =  $4.0 \text{ mm}^2 \pm 0.523$  [SD], experimental =  $2.26 \text{ mm}^2 \pm 0.77$ ;  $p < .05$ ). The smaller forelimb region of experimental animals resulted from a reduction in the size of the rostral forelimb zone and an absence of the forelimb portion of the OLZ. We used electrophysiological recording methods to identify somatic sensory receptive fields in the area where the OLZ is normally found. Unlike the distal cutaneous receptive fields normally present in

the OLZ (8, 10) cells in this region in experimental rats ( $n = 3$ ) were activated by cutaneous inputs from the trunk and shoulder regions. This cutaneous representation formed part of an enlarged SI territory for these body parts.

By contrast, agranular cortex in experimental rats did not contain large inexcitable areas; ICMS at most sites activated muscles at low thresholds. Uncharacteristic of the normally high threshold for proximal forelimb movements in normal rats, we found many sites in amputated animals in which shoulder movements were evoked at the lowest threshold. These thresholds were often comparable to those necessary for activation of more distal forelimb muscles in intact animals and were lower than shoulder movement thresholds in normals (control threshold [ $n = 28$ ] =  $50.07 \mu\text{A} \pm 12.5$ ; experimental [ $n = 78$ ] =  $37.99 \mu\text{A} \pm 14.46$ ;  $p \leq .0005$ ). This finding suggests that neonatal forelimb nerve injury increased the strength of the connection between MI and the proximal muscles.

A significant feature of the organization of MI in the normal rat is the distinct segregation of body parts. Thus, although stimulation at a single site will often activate muscles of, for example, the distal and proximal forelimb (Fig. 1C), it is rare to find the forelimb and vibrissa, as another example, represented at the same site. The only exception to this appears to be at borders between representations for two body parts. However, in experimental animals (Fig. 1D) the shoulder and vibrissa frequently shared common sites in the forelimb region. Frequently vibrissa movements were evoked at lower stimulation currents than activation of the shoulder musculature. This intermingling of representations indicates that the vibrissa representation expanded into the presumptive forelimb zone, while the relationship of MI with proximal forelimb muscles was preserved within this same area. A possible mechanism of the expanded MI vibrissa representation could be a shift of corticospinal projections from the cervical cord to brainstem targets that control vibrissa movements. However, injections of HRP into the cervical spinal cord (15) showed a similar distribution of corticospinal neurons in normal and experimental rats. Additionally, MI cytoarchitecture lacked any obvious change that might suggest anatomical reorganization.

In contrast to the integrity of the general aspects of MI cytoarchitecture and corticospinal projections, forelimb removal at birth resulted in marked anatomical changes within the spinal cord and the primary afferent pathway. Sections through the spinal cord showed a near total absence of large, darkly staining cells where distal forelimb motor neuron pools are normally found; these pools appeared to be normal on the side of the intact limb (16). There was an obvious shrinkage of the ipsilateral dorsal columns and a loss of cells in the ipsilateral cuneate nucleus.

These experiments indicate that environmental factors during development influence the pattern of movement representation in motor cortex. We conclude that forelimb removal in neonatal rats results in three forms of change in MI representation patterns identified by ICMS mapping. First the MI has more effective or stronger connections with proximal muscles compared to normal animals. Second, cortically adjacent representations expand into the presumptive forelimb territory and third, in the absence of normal target structures, the region of the cutaneous part of the MI forelimb area, the OLZ, fails to develop. However, cutaneous input to the OLZ is preserved through the expansion of the SI trunk and shoulder representations.

The basis for MI reorganization is uncertain though several different mechanisms might be involved. The threshold change for proximal body parts could represent a strengthening of already existing synaptic connections between MI and alpha motor neurons for the shoulder musculature. Alternatively, new connections between MI neurons and their target cells or between MI target cells (such as interneurons) and their targets may have been formed. Determination of reorganization sites will require mapping of these subcortical structures. The "expansion" of other body parts into the presumptive forelimb region may also result from changes in functional connectivity between MI and its targets. However, a more likely explanation for the expanded vibrissa representation is that forelimb removal at birth preserves widespread connections that are transiently formed during normal development and then lost (5, 17, 18). Thus, individual cells in MI may project subcortically to vibrissa and to proximal and distal forelimb motor control structures early in development and then be restricted largely to one target as the organism matures. This hypothesis requires testing. Our anatomical finding that corticospinal cell distribution was not

altered by neonatal forelimb amputation may suggest that some cells in the mixed vibrissa-shoulder area retain weak or ineffective projections to the spinal cord.

The absence of the OLZ in MI may require a different explanation than retention of developing pathways. Functionally the OLZ is a mixed area, responding to cutaneous inputs much like other regions of SI, while also contributing directly to motor output, as demonstrated by microstimulation induced movement. It is possible that as a result of the close coupling between the sensory and motor functions in the OLZ, depriving this area of its normal complement of peripheral somesthetic inputs inhibits the formation of normal efferent connections. MI in the rat is not responsive to cutaneous stimulation (8-10, 19), so a similar mechanism could not operate in the observed alterations in MI output organization, but aberrations in other inputs could induce change. Alternatively, damage to motor nerves could form a basis for motor system reorganization.

Both lesions of peripheral receptors and sensory deprivation are known to modify patterns of cortical sensory representation (2). These changes result from reorganization of input pathways at the cortical level, and in many cases at the subcortical levels as well (3, 20). Our results indicate that cortical output organization is also subject to environmental influences. It appears that a general principle of the modifiability of cerebral cortex, whether it is related to sensation or motor output, is emerging. An implication of these results is that MI and perhaps the entire cortex may be undergoing continual reorganization. In general, cortical modifiability may depend on the availability of target cells, inputs from other brain regions and inputs from peripheral end organs. An example of this suggestion can be found in a clinical study in which the somatic sensory evoked potentials of human amputees changed over time (21). Another example would be in cases of cerebral hemorrhagic infarcts when humans, who often are initially paralyzed, recover varying degrees of motor function. The mechanisms for such recovery are uncertain, but they could involve changes in the somatotopy of MI.

## References and Notes

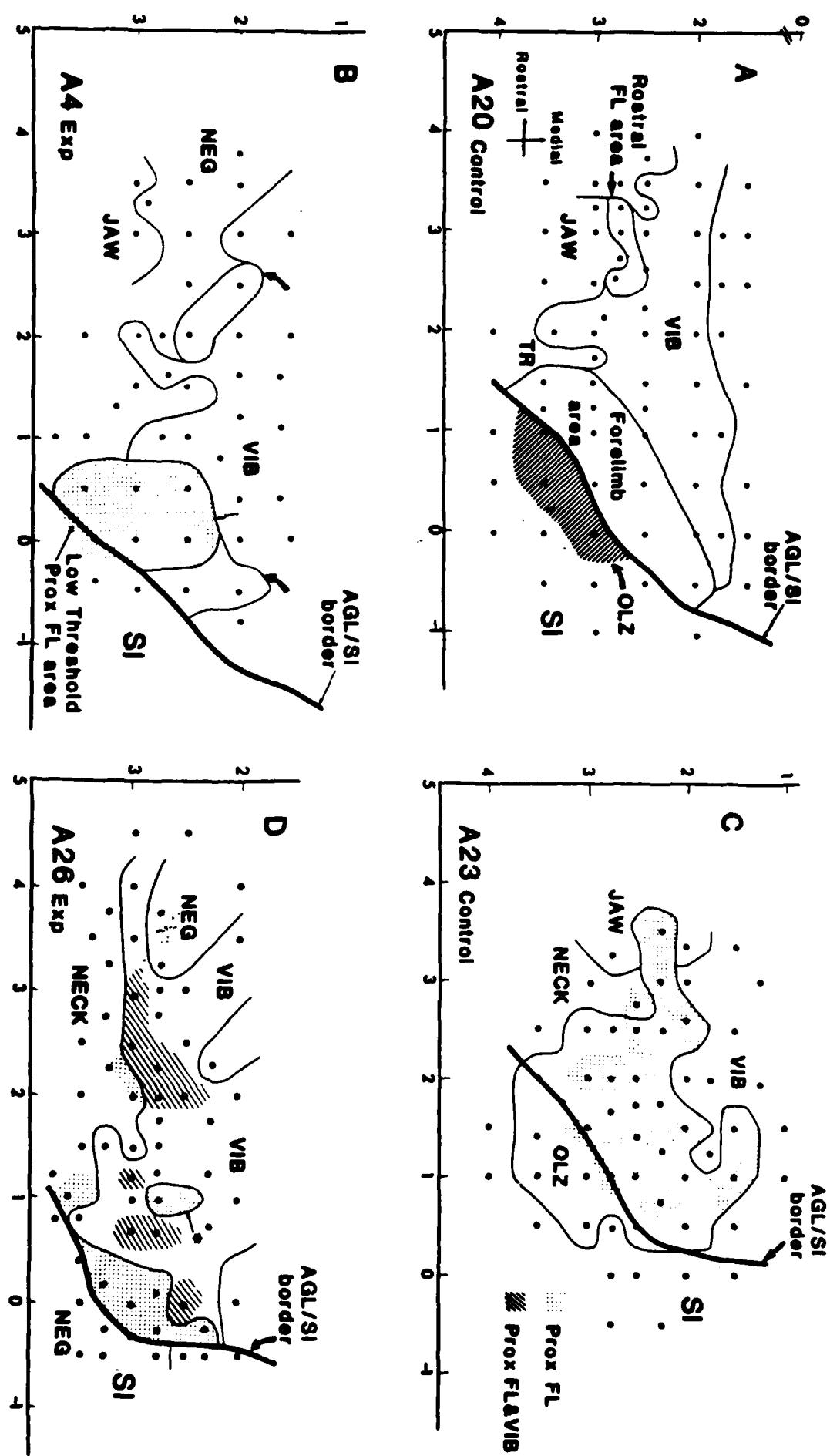
1. E. V. Evarts, In: Handbook of Physiology, Section I, the Nervous System, Vol. II, Motor Control, V. B. Brooks, Ed. (American Physiol. Soc., Bethesda, MD, 1981), pp. 1083-1120; C. G. Phillips and R. Porter, Corticospinal Neurones, (Academic Press, London, 1977).
2. J. H. Kaas, M. M. Merzenich, H. P. Killackey, Ann. Rev. Neurosci. **6**, 325 (1983); M. M. Merzenich *et al.*, Neurosci. **8**, 33 (1983); \_\_\_\_\_, ibid. **10**, 639 (1983); T. N. Wiesel and D. H. Hubel, J. Neurophysiol. **28**, 1029 (1965).
3. C. Shatz and M. Stryker, J. Physiol. (Lond) **281**, 267 (1978); D. H. Hubel, T. N. Wiesel, S. LeVay, Phil. Trans. Royal Soc., Lond. Series B **278**, 377 (1977).
4. G. O. Ivy and H. P. Killackey, J. Comp. Neurol. **195**, 367 (1981); \_\_\_\_\_, J. Neurosci. **2**, 735 (1982).
5. G. M. Innocenti and D. O. Frost, Exp. Brain Res. **39**, 365 (1980).
6. C. N. Woolsey, In: The Biological and Biochemical Bases of Behavior, H. F. Harlow and C. N. Woolsey, Eds. (Univ. of Wisconsin Press, Madison, 1958), pp. 63-81.
7. Detailed maps of representation patterns in MI have been elaborated by ICMS. With this technique electrical currents in the microampere range are applied through a microelectrode inserted among cortical output cells in layer V to identify the muscles or movements that are activated from that cortical site; S. D. Stoney, Jr., W. D. Thompson, H. Asanuma, J. Neurophysiol. **31**, 659 (1968). Although ICMS produces artificial patterns of neural activation, it appears to reflect the functional relationship of small regions of MI with muscles. Thus, neurons at a cortical site discharge during movements of the same muscles that are activated by ICMS at that same site; D. R. Humphrey and D. J. Reed, In: Motor Control Mechanisms in Health and Disease, Advances in Neurology, J. E. Desmedt, Ed. (Raven Press, New York, 1983), vol. 39, pp. 347-372.
8. R. D. Hall and E. P. Lindholm, Brain Res. **66**, 23 (1974).
9. Donoghue, J. P. and S. P. Wise, J. Comp. Neurol. **212**, 76 (1982).
10. K. J. Sanderson, W. I. Welker, G. M. Shambes, Brain Res. **292**, 251 (1984).

11. The OLZ also has connectional features characteristic of SI and MI; J. P. Donoghue, K. L. Kerman, F. F. Ebner, J. Comp. Neur. **183**, 647 (1979); S. P. Wise and J. P. Donoghue, In: The Cerebral Cortex, The Functional Areas of the Cerebral Cortex, E. G. Jones and A. Peters. Eds. (Plenum, New York, 1986) vol. 5, p. 243-270.
12. In rat pups less than 24 hours old, the right forelimb was removed surgically at the shoulder joint during hypothermic anesthesia. The wound was sutured closed and the pups were returned to the dam. Mapping was carried out at 2-4 months of age.
13. The acute mapping procedures have been described previously (9). Briefly, animals were anesthetized with ketamine hydrochloride (intraperitoneal injections) and mounted in a stereotaxic frame. Microelectrodes (PtIr, with 5  $\mu$ m tips, 0.5-2 M $\Omega$  impedance at 1 KHz) were lowered to a depth of  $1.8 \pm 0.1$  mm below the pial surface. Current trains (30 msec duration, 300 Hz, 200  $\mu$ sec monopolar cathodal pulses) of 5-60  $\mu$ A were routinely passed through the electrode tip while we examined the body visually and by touch to determine which body parts moved or muscles contracted. The resolution of the electrode penetrations was between 0.1-0.5 mm. At the termination of each experiment lesions were made at selected sites by passing a DC current of 10  $\mu$ A for 10 sec. The animals were perfused, the brains removed and processed for Nissl substance or cytochrome oxidase staining. In reconstructions, map borders were defined as the mid-point between two distinct body parts.
14. E. J. Neafsey and C. Sievert, Brain Res. **232**, 151 (1982).
15. The corticospinal tract of 4 normal and 4 experimental rats was injected at the C4-5 level with 0.5  $\mu$ l of 30% HRP using a 10  $\mu$ l Hamilton Syringe. Rats survived 2 days. They were perfused with mixed aldehydes and tangential sections of the flattened cerebral cortex were reacted for HRP with TMB as the chromagen, according to standard techniques described previously; J. P. Donoghue and C. Parham, J. Comp. Neurol. **217**, 390 (1983). Cell distributions were charted on outline drawings of sections using a drawing tube attached to a microscope.

16. Sections of formalin fixed brains and spinal cord were cut in the frontal plane on a freezing microtome, stained with thionin using routine procedures and were compared microscopically.
17. S. S. Easter, Jr., D. Purves, P. Rakic, N.C. Spitzer Science **230**, 507 (1985); G. M. Innocenti and D. O. Frost, Nature **280**, 231 (1979); T. Cabana and G. F. Martin, Develop. Brain Res. **23**, 69 (1985).
18. D. D. M. O'Leary, B. B. Stanfield, W. M. Cowan, ibid. **1**, 607 (1981).
19. J. P. Donoghue, Brain Res. **333**, 173 (1985).
20. M. Devor and P. D. Wall, Nature **276**, 75 (1978); \_\_\_\_\_, J. Comp. Neurol. **199**, 277 (1981).
21. R. E. P. Sica, O. P. Sánz, L. G. Cohen, J. D. Freyre, M. Panizza, Electromyogr. clin. Neurophysiol. **24**, 415 (1984).
22. We thank B. Z. Berger, B. Schlaggar, M. McGavic and S. Ryan for their assistance in the data collection and anatomical analyses and Dr. James C. Houk for commenting on an earlier version of this manuscript. This work was supported by NIH grant # 1 R01 NS22517-01 and was also supported in part by the U.S. Office of Naval Research contract # N00014-81-K-0136.

### Figure Legends

**Fig. 1. Microstimulation maps of forelimb region of MI in two control and two experimental rats.** In the four panels, each dot marks a stimulation site and thin solid lines mark boundaries between somatotopic subdivisions of MI. The border between the frontal agranular and granular SI cortex is marked by a heavy line. Numbers on axes are in millimeters, with bregma as the zero point. (A) Pattern of representation at the lowest current intensities showing the location of the forelimb representation in MI. The forelimb is divided into two regions; the caudal section ("Forelimb area") is contained largely in frontal agranular cortex (AGL) but also extends into the adjacent granular SI cortex (OLZ). A second forelimb region ("rostral FL area") is also contained within frontal agranular cortex. Stimulation at any site within the total extent of the forelimb region produces distal forelimb movements. (B) Low threshold map from an experimental rat. Movements of contralateral shoulder musculature (stippled) and of the ipsilateral forelimb (enclosed areas marked by curved arrows) are now evoked at the lowest currents. The presumptive forelimb area and MI does not overlap with SI. In normal rats ipsilateral sites coincide with some contralateral distal forelimb sites (unpublished observations, Donoghue and Schlaggar). (C) Coincidence of distal forelimb and shoulder sites in a normal rat. Shoulder movements (stippled area) are found at higher thresholds at some sites within the distal forelimb area. One exception is seen at the border with the neck area. (D) The abnormal coincidence of vibrissa and shoulder points in an experimental rats. At each shaded site vibrissa movements were elicited at the lowest threshold, but at higher current intensities shoulder movements were evoked.



END

DTIC

7-86